


ssDRIP-seq and DRIP-qPCR

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R-loops coordinate with SOX2 in regulating reprogramming to pluripotency

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Detailed protocol

gDNA extraction

Resuspend the pellet in TE buffer (1.6–6.4 ml, depending on the quantity of pellet) and add SDS (final concentration: 0.5%) and Proteinase K (final concentration: 0.1 mg/ml). Then the mixture was incubated at 37°C in a shaker with 180–240 rpm for 4–12 hours.

Add 1/4 volume 5 M KAc, mix and place the tube on ice, 10–20 min.

Add 1 volume phenol:chloroform:isoamyl alcohol (25:24:1), mix well by Vortex, and centrifuge at 12000 g for 10 min at 4°C.

Transfer the supernatant to a new tube, add 1 volume chloroform:isoamyl alcohol (24:1), and mix well by vortex. Centrifuge at 12000 g for 10 min at 4°C.

Transfer the supernatant to a new tube, and centrifuge again at 12000 g for 5 min at 4°C. Transfer only 90% of the supernatant.

Precipitate the supernatant by adding 0.7 volume isopropanol, and incubate for 30 min at room temperature. Centrifuge at 12000 g for 10 min at 4°C and wash the pellet with 1 ml 70% ethanol. Air dry the pellet and resuspend with 200–500 µl ddH₂O.

Cocktail of restrict enzymes digestion

Digest 2–10 µg gDNA in 200 µl final volume, 8–16 hours, by RE combinations. MseI, DdeI, AluI, and MboI (final concentration: 100 U/ml for each enzyme). Incubate at 37°C overnight.

Add 1 volume phenol:chloroform:isoamyl alcohol (25:24:1), mix well by Vortex, and centrifuge at 12000 g for 10 min at 4°C.

Transfer the supernatant to a new tube, add 1 volume chloroform:isoamyl alcohol (24:1), and mix well by vortex. Centrifuge at 12000 g for 10 min at 4°C.

Transfer the supernatant to a new tube, and centrifuge again at 12000 g for 5 min at 4°C. Transfer only 90% of the supernatant.

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Fragmented gDNA was quantified by Qubit.

DRIP

Take 10 µg fragmented gDNA to a 1.5 ml Safe-lock EP tube and add 50 µl 10X DRIP binding buffer (100mM NaPO₄ pH7.0, 1.4M NaCl, 0.5% Triton X-100), and 10 µg S9.6 antibodies. (store the rest gDNA as input DNA for input-library or input of qPCR).

Incubate for 8–16 hours at 4°C while gently inverting on a rotisserie shaker.

50 µl Dynabeads (Protein A or G) are used for each tube. Beads are pre-washed with 1X DRIP binding buffer (10x diluted in TE) for 3 time, and each wash take 5–10 min with gentle shaking at 4°C.

Add DNA/S9.6 complexes to pre-washed beads, and incubate with gentle shaking at 4°C for 3–4 hours.

Wash 4 times beads/antibody complexes (remove background) with 1X DRIP binding buffer at room temperature, and each wash takes 5–10 min.

Add 250 µl elution buffer (50mM Tris pH8.0, 10mM EDTA) and 200 µg Proteinase K (10 µl) to beads/antibody complexes, incubate for 60 min in an Eppendorf ThermoMixer at 55°C, 1000 RPM.

Cleanup elution with 250 µl phenol:chloroform extraction, and move supernatant to a new tube. Then add 1/10 volume 3 M NaAc, 1 µl glycogen and 2.5 volume ethanol. Precipitate the DNA at -20°C for 2 hours.

ssDRIP-seq library was constructed by using the Accel-NGS 1S Plus DNA Library Kit (Swift Biosciences).

How to cite: (Readers should cite both the Bio-protocol preprint and the original research article where this protocol was used)

- Li, Y., Xu, W., Song, Y., Li, Q., Sun, Q. and Yao, H. (2020). ssDRIP-seq and DRIP-qPCR. Bio-protocol Preprint. bio-protocol.org/prep597.
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